cellArtgen

Regenix™ Xenograft
PROTOCOL

Regenix™ Xenograft is designed for patient-derived (PDX) and cell line-derived (CDX) xenograft applications. Composed of various basement membrane proteins from normal porcine tissues and featuring atelocollagen for biological safety, it offers easy sample preparation and injection. Regenix™ Xenograft ensures high cell retention, promoting rapid tumor growth with a smooth, round shape, making it a reliable tool for consistent, high-quality research results.

PROCEDURE

CDX model generation using Regenix™ Xenograft

01 Thaw Regenix™ Xenograft for at least 4 hours by submerging the vial in an ice bucket and storing it in a 4°C refrigerator before use. Avoid multiple freeze/thaw cycles.

02 Cut the tip off a 200 μL pipette tip with sterile scissors to obtain an opening diameter of 1.5-2 mm, and mix Regenix™ Xenograft by slowly pipetting; Be careful not to create air bubbles during this process.

> **Note** Regenix™ Xenograft may have high viscosity, so there may be some difficultly in pipetting. If a lot of bubbles are generated after pipetting, centrifuge before use.

> **Note** Regenix™ Xenograft may form a gel at temperature above 10°C. The temperature must be lowered to between 4°C and 8°C throughout all handling processes to ensure depolymerization.

03 Add Regenix™ Xenograft to the cell pellet and resuspend evenly by slow pipetting.

> **Note** Keep the mixture cold to prevent the Regenix™ Xenograft from solidifying before injection.

04 Using a chilled syringe with a 25-29 gauge needle, draw up the 100 μL cell-Regenix™ Xenograft mixture.

05 Inject the cell-Regenix™ Xenograft mixture subcutaneously into the flask of the anesthetized mouse. Slowly administer the entire volume of the mixture.

> **Note** Insert the needle at a shallow angle to ensure the injection remains in the subcutaneous space and does not penetrate the muscle.